

# **#PF213-0.25-EX** For 250 µL Reaction

in vitro research use only Store at -80°C before opening

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## Kit components

<ul> <li>Solution I<sup>*1</sup> Amino acids, NTPs, tRNAs and subs Store at -20℃</li> </ul>	<b>100 μL</b> strates of enzymes
• Solution II *2 Proteins in 30% glycerol buffer Store at -20°C or -80°C *3	12.5 μL
• Solution III <sup>*2</sup> Ribosomes (20 μM) Store at -80°C <sup>*3</sup>	25 μL
• Cysteine <sup>*1</sup> Cysteine (10 mM) Store at -20°C	20 µL
• DTT *1 Dithiothreitol (40 mM) Store at -20°C	20 µL
• <b>GSH</b> <sup>*1</sup> Reduced glutathione (80 mM) Store at -20°C	20 µL
• DHFR DNA <sup>*2, 4</sup>	10 uL

### PCR product (20 ng/µL) containing a gene encoding E. coli DHFR Store at -20°C

## Introduction

### About PURE frex®

PUREfrex<sup>®</sup> is a reconstituted cell-free protein synthesis kit which GeneFrontier has developed based on the PURE system technology. The target protein can be synthesized by adding the template DNA (or mRNA) to the reaction mixture. The PURE system is a unique cell-free protein synthesis system, which has originally developed by Professor Takuya Ueda at the University of Tokyo, and consists of only purified factors necessary for transcription, translation and energy regeneration (Ref. 1). Therefore it enables to adjust the composition of the reaction mixture.

PURE frex<sup>®</sup> has been raised in purity by improving the methods for preparing ribosomes, tRNAs and all proteins in the reaction mixture compared with the original PURE system (Ref. 2). As the result, the contaminating lipopolysaccharide from E. coli is reduced to around 0.1 EU per 1 µL of reaction and other contaminants, such as RNase and β-galactosidase, are also reduced.

Because all of proteins in PURE frex® have no tags, the synthesized protein can be purified and detected by any tags.

#### (References)

1. Shimizu et al. (2001) Nat. Biotechnol., vol. 19, p. 751. 2. Shimizu et al. (2005) Methods, vol. 36, p. 299.

## Kit components

#### Store at -80 °C before opening

### \*1)

Reducing agents are excluded from Solution I in PUREfrex<sup>®</sup> 2.1. Please add attached Cysteine and reducing agent (DTT or GSH) to the reaction mixture.

The standard working concentration of Cysteine, DTT and GSH are 0.5, 2 and 4 mM, respectively. Other reducing agents are also available for the reaction, e.g. 2-mercaptoethanol and TCEP.

We recommend optimizing the concentration of reducing agent to synthesize a protein with higher activity because the suitable concentration is depends on the synthesized protein.

#### \*2)

Solution II, Solution III and DHFR DNA are the same as PUREfrex® 2.0 (#PF201).

#### \*3)

For storage at -80°C, the remaining solution should be frozen rapidly in liquid nitrogen or dry ice/ethanol. Please divide into aliquots, if necessary, and avoid refreeze and thaw as much as possible.

#### \*4)

As a positive control for the protein synthesis reaction, 1  $\mu$ L of DHFR DNA should be added to 20 µL of reaction. Please visit our web site to get the nucleic acid sequence of DHFR DNA.

## **Template DNA**

### 1. Construction of the template DNA



(Ribosome Binding Site; SD sequence)

The template DNA for the protein synthesis by PUREfrex<sup>®</sup> should contain T7 promoter and ribosome binding site (SD sequence) at the upstream of the gene encoding the target protein.

At the downstream of stop codon, more than 10 nucleotides are required.

All three stop codons are available.

Both circular and linear DNAs are available as a template DNA.

For the circular DNA, e.g. plasmid DNA, T7 terminator is required at the downstream of stop codon.

For the linear DNA, e.g. PCR product or DNA fragment digested by restriction enzymes, T7 terminator is not necessarily required.

## Protocol

The protein synthesis reaction using PUREfrex® 2.1 can be performed in any volume. For example, 20 µL of reaction mixture containing 0.5 mM Cysteine and 4 mM GSH is assembled as below.

- 1. Thaw completely Solution I, Cysteine and GSH by incubation at room temperature or 37 °C for 1 minute. and then cool on ice.
- 2. Thaw Solution II and Solution III on ice.
- 3. Mix each solution by vortex and centrifuge briefly to collect solution at the bottom.
- 4. Assemble the reaction mixture in a tube as follows. (Add the template DNA to 0.5-3 ng/µL per 1 kbp)

Water	7 -X μL
Solution I	8 μL <sup>*5)</sup>
Cysteine (10 mM)	1 μL
GSH (80 mM)	1 μL
Solution II	1 μL
Solution III	2 µL
Template DNA	XμL
Total	20 µL

5. Incubate the tube at 37°C for 2-6 hours. 6. Analyze the synthesized product.

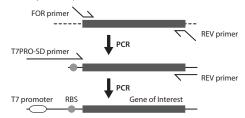
#### \*5)

Please note that the volume of Solution I in PUREfrex® 2.1 (#PF213) is different from PUREfrex® 2.0 (#PF201).

# **Template DNA**

### 2. Prepatation of the template DNA

An example of the method for preparing the template DNA by two-step PCR is shown in below.



### 3. Sequence of primers

#### FOR primer

5'-AAGGAGATATACCA-ATG-N(10-20)-3' RBS

**REV** primer

5'-GGATTAGTTATTCA-TTA-N(10-20)-3' more than 10 any nucleotides

T7PRO-SD primer

5' - GAAATTAATACGACTCACTATAGGGAGACCACAACGGTTTCCC T7 promoter TCTAGAAATAATTTTGTTTAACTTTAAGAAGGAGATATACCA-3' RBS

### Note

PUREfrex® is developed for in vitro research use only. PUREfrex® should not be used for therapy, diagnostic or administration to animals including human and should not be used as food or cosmetics etc.

To avoid the contamination of nuclease, nuclease-free water, reagents and equipment should be used. We also recommend wearing gloves and mask.

For information concerning commercial use of PUREfrex®, please contact GeneFrontier. e-mail:purefrex@genefrontier.com

> ھ GeneFrontier Corporation www.genefrontier.com

## Distributor

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